Orginal article

Caffeine (1,3,7-Trimethylxanthine) as a modulator of Arsenic bioaccumulation in the experimental *Pelecypod* model, *Lamellidens corrianus*

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ABSTRACT

Aims: The present investigation was carried out to study modulator effect of caffeine (1,3,7-Trimethylxanthine) on Arsenic-induced alterations on freshwater bivalve, *Lamellidens corrianus*.

Materials and Methods: Freshwater *Pelecypod Mollusc, L. corrianus* were exposed to acute dose of arsenic (0.672 ppm As³⁺) for 4 days. Arsenic exposed *Bivalves* were allowed to cure naturally and with caffeine. Testis and digestive glands from different groups were removed and dried in the oven. The dry powders were digested in nitric acid and perchloric acid in 4:1 ratio at hot temperature till dryness. The digest was dissolved in double glass distilled water and the arsenic contents were estimated by the Atomic Absorption Spectrophotometer.

Results: After 4 days of exposure, the amount of bioaccumulated arsenic in the testis and digestive glands was 1.044 and 1.119 μ g/g. During the recovery after 4 days, the bioaccumulated arsenic was reduced to 0.969 and 1.067 μ g/g of normal water while in caffeine exposed *Bivalves* was reduced to 0.917 and 0.975 μ g/g in testis and digestive glands respectively.

Conclusion: Rapid reduction in the arsenic contents in caffeine exposed *Bivalves* indicates the role of caffeine in arsenic excretion. Therefore, the caffeine (1,3,7-Trimethylxanthine) as a modulator of Arsenic bioaccumulation in the experimental *Pelecypod* model, *L. corrianus*

Key words: Arsenic, Bioaccumulation, Caffeine, Lamellidens corrianus

INTRODUCTION

The *Bivalves* being bottom dwellers are exposed to higher doses of the toxic metals as most metal precipitates and increases in the sediments.^[1,2] The high metal contents in the surrounding make them the best bioaccumulators.^[3,4] These mussels are useful indicators of the abundance and spatial distribution of metals in aquatic ecosystem.^[5-7]

Investigation results those acute disorders in aquatic organisms due to toxic heavy metals.^[8] Heavy metals are very

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harmful pollutants as they remain in the nature for a very long period. The bioaccumulation of heavy metal toxicants in aquatic animals depends on availability and persistence of the contaminants in water and the physico-chemical properties of toxicants. The metal content of mollusk is controlled by the surface to volume ratio of the animal.^[9,10]

Arsenic is a chemical that bioaccumulates in tissues of aquatic organisms but does not biomagnify in the aquatic food chain.^[11-16] The consensus in the literature is that 85-90 percent of arsenic found in edible portions of marine fish and shellfish is organic arsenic Arsenobetaine (ASB), Arsenocholine, Dimethylarsinic Acid, Monomethylarsonic Acid (MMA) and that appropriately 10 percent is inorganic arsenic.^[17-20] Less is known about the forms of arsenic in freshwater fish, but there is evidence that organic arsenic may be prevalent, Field based study or considerably less.^[14,21-25]

More recent research indicates that as compared to arsenite, trivalent methylated arsenic metabolites exerts a number of unique biological effects, are more cytotoxic and genotoxic, and are more potent inhibitors of the activities of some enzymes.^[26,27] Because each arsenic species (As^{III}, As^V, ASB, MMA^V, and MMA^{III}) exhibits different toxicities, it may be important to take into account the fraction of total arsenic present in the inorganic arsenic and organic arsenic forms while estimating the potential risk posed to human health through the consumption of arsenic contaminated fish and shellfish.^[13]

The removal of toxic metal through chelating allows the body functions at an optimal level. Dissolved heavy metal ions are positively charged, caffeine contains uncharged or negatively charged molecules, and the metal ions can be taken out of solution by binding to negatively charged molecules in the coffee which indicates that caffeine can have the capacity to bind the heavy metals from living organisms. According to investigator, oxygen from 2nd and 6th position of caffeine forms chelate with heavy metal.^[28] Therefore, it is proposed to study the efficacy of caffeine in the detoxification of arsenic with respect to their tendency of bioaccumulation in an experimental animal model, the freshwater bivalve, *Lamellidens corrianus*.

MATERIALS AND METHODS

L. corrianus collected from the Nathsagar Dam, Paithan were acclimatized in the laboratory condition at room temperature for 5 days. The healthy and active acclimatized *Bivalves* of approximately the same size were divided into two groups A and B.

- 1. A group *Bivalves* were maintained as a control,
- B group *Bivalves* were exposed to acute doses (LC₅₀) of sodium arsenate (0.672 ppm As³⁺). After 4 days *Bivalves* from group B were divided into two groups C and D.
- 3. C group *Bivalves* pre-exposed to acute dose of sodium arsenate were allowed to cure in normal dechlorinated water.
- 4. D group *Bivalves* pre-exposed to acute doses of sodium arsenate were exposed to 5 mg caffeine⁻¹ in dechlorinated water.

The experimental *Bivalves* of A and B group were dissected after 24 h and 96 h and from C and D groups of recovery after 2 days and 4 days. Testis and digestive glands of bivalves from all groups were dried at 80°C in an oven till constant weight was obtained. The powders obtained were stored in airtight specimen bottles by waxing the cork outside.

The 500 mg tissue powder was digested in 10 ml of acid mixture (HNO₃: Perchloric acid) in (4:1) ratio on a hot plate till dryness. The digested mixture was cooled and diluted to 50 ml by double distilled glass water in a volumetric flask and was filtered by (Whatman grade 541) filter paper. From each sample, arsenic was estimated by Atomic Absorption Spectrophotometer (Chemito)

The concentration of arsenic accumulated in the tissue of each exposure period in each tissue was recorded and the results are given in the Table 1 and Figure 1a and b.

| Treatment | | Tissue | Exposure period | | Recovery Period | |
|--|--------------------------------------|---------------------|-------------------|-------------------|---------------------------------------|--|
| | | | 24 h | 96 h | 2days | 4days |
| Control (A) | | Testis | 0.016 ± 0.006 | 0.016 ± 0.002 | _ | _ |
| | | Digestive Glands | 0.011 ± 0.004 | 0.011 ± 0.003 | _ | _ |
| 0.672 ppm As ³⁺ (B) | | Testis | 1.026 ± 0.051 | 1.044 ± 0.037 | _ | |
| | | Digestive Glands | 1.089 ± 0.023 | 1.119 ± 0.016 | _ | _ |
| After 96 h Exposure to 0.672 ppm As ³⁺ | Normal Water (C) | Testis | _ | _ | 1.010±0.021 [-3.256] | 0.969±0.012 [-7.183] |
| | | Digestive Glands | _ | — | 1.105±0.014 [-1.251] | $\begin{array}{c} 1.067 \pm 0.024 \\ [-4.647] \end{array}$ |
| | Normal Water + 5mg/l Caffeine (D) | Testis | _ | _ | 0.942±0.036 [-9.770] (- 6.732) | 0.917±0.052 [-12.167] (-5.366) |
| | | Digestive Glands | _ | _ | 1.073±0.028 [-4.110] (-2.895) | 0.975±0.011 [-12.868] (-10.032) |

Table 1: Arsenic content (μ g/g dry weight) in testis and digestive glands of *Lamellidens corrianus* after acute exposure to As³⁺and during recovery

Values in the [] brackets indicate percent change over respective B of 96 h Values in the () brackets indicate percent change over respective C

Shamsundar and Sureshchandra: Caffeine as a Modulator on Arsenic Bioaccumulation in L. corrianus

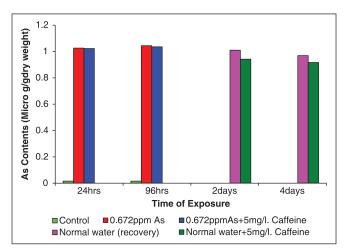


Figure 1a: Arsenic contents in testis of *L. corrrianus* after acute treatment and recovery

RESULTS

Bioaccumulation of arsenic in testis and digestive glands of *L. corrianus* exposed to arsenic (0.672 ppm As^{3+}) and during recovery has been summarized in the Table 1 and Figure 1a and b.

Bioaccumulation of arsenic

Testis

Bivalves from control group showed 0.016 μ g/g arsenic in testis while the amount of accumulated as in presence of sodium arsenate (0.672 ppm As³⁺) for 24 h was 1.026 μ g/g and after 96 h was 1.044 μ g/g. The Bivalves pre-exposed to as showed fast arsenic depletion with caffeine than those allowed to cure naturally. The amount of accumulated as observed after 2 days was 0.942 μ g/g and after 4 days was 0.917 μ g/g while in those Bivalves allowed to cure naturally, the amount of accumulated arsenic after 2 days was 1.010 μ g/g and after 4 days was 0.969 μ g/g.

Digestive glands

The control group of animals showed 0.011 μ g/g arsenic in digestive gland. The amount of bioaccumulated as in presence of sodium arsenate (0.672 ppm As³⁺) for 24 h was 1.089 μ g/g and after 96 h was 1.119 μ g/g. The *Bivalves* preexposed to sodium arsenate showed fast removal of As with Caffeine than those allowed to cure naturally. The amount of accumulated As observed after 2 days was 1.073 μ g/g and after 4 days was 0.975 μ g/g in caffeine exposed *Bivalves* and in those allowed to cure naturally, the amount of accumulated As³⁺for 2 day was 1.105 μ g/g and 4 days was 1.067 μ g/g.

DISCUSSION

Heavy metal uptake and concentration in the food chain, especially those terminating in the human beings have renewed interest largely due to several instances

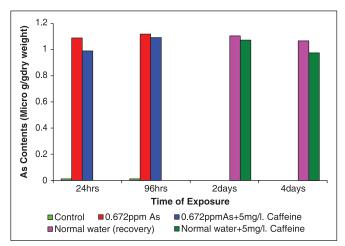


Figure 1b: Arsenic contents in digestive glands of *L. corrianus* after acute treatment and recovery

of human intoxication.^[29,30] The accumulation of metal in different species is the function of their respective membrane permeability and enzyme system.^[31] Toxicity and bioaccumulation of arsenic and chromium in epigen and hypogen in freshwater macro-invertebrates tolerates, and higher values of Zn, Ni, Hg, Cr and Pb were observed in comparison to water in fish.^[32,33]

Higher amount of As, Hg, Mn, Ni, Cr, Cd, Pb, Fe, Cu and Zn in the *Bivalves*, *Ruditapes decussatus* and *Ruditapes philippinarum* from the Atlantic coast of Southern Spain observed.^[34] *Bivalves*, *Pinctadea radiata* and *Brachidontes pharaonis* were bioaccumulate fairly large amount of Cd, Fe, Zn, Cu.^[35]

The ratio between bioaccumulation and exposure concentration with periods of exposure has been shown by various investigators.^[36,40] The accumulation of several metals is due to the low capacity of these mollusks for discriminating among metals, which are similar in some characteristics such as ionic radius.^[41,42] Mussels also possess a variety of effective detoxification mechanisms to reduce the toxicity of the metal uptake.^[43,45]

Caffeine binds divalent cations of calcium, which indicates that caffeine can bind other reactive divalent cations of heavy metals. Calcium association with caffeine is an indicator of such binding.^[46]

Suppressive effect of caffeine on hepatitis and apoptosis induced by tumor necrosis factor L studied.^[47] The interactions between caffeine and metal ions can be through its oxygen and nitrogen atoms, Because of the blockage on N1, N3 and N7 atoms by methyl groups. Caffeine probably binds to metal ions through its oxygen atom of 2nd and 6th position. Caffeine would bind to metal ions through its second and sixth O atoms in the gaseous phase.^[48]

Caffeine inhibited hepatocarcinogenesis induced by 2-acetylaminoflure showed.^[49] Oral administration of

green tea and caffeine inhibits Ultra Violet-induced carcinogenesis.^[50]

The binding capacities of caffeine with different micronutrients is weaker than that of the EDTA (Ethylene Diamine Tetra Acetic acid) and since all nitrogen groups are blocked by methylation, metals probably form complexes with second and sixth oxygen of caffeine.^[28] Though the binding strength of caffeine with metal ions is weaker than that of EDTA, it is sufficient to drag the metal ions bound to the proteins because the metals are usually bound to –SH groups of proteins and the bond energy of the oxygen metal complex is stronger than sulphydril-metal bond. EDTA is very toxic as compared to the caffeine, and it removes essential micronutrients from the body.

The kidney manufacture and discharge up to 100% more urine on caffeine intake, investigation state the increased urinary excretion of calcium, magnesium, sodium and chloride after oral doses of caffeine.^[51] The caffeine metalchelate complex being of small molecular weight can easily pass through the membranes of the cells of the epithelia of the tubules of the kidney and can be effectively excreted out.

The bioaccumulation of arsenic in *L. corrianus* was studied. The arsenic accumulation increased with the increase in exposure period of acute concentration of arsenic (0.672 ppm As³⁺). The amount of arsenic accumulation is less when exposed with 5 mg/l caffeine in presence of respective concentration of arsenic as compared to those exposed to only arsenic. The *L. corrianus* pre-exposed to arsenic (0.672 ppm As³⁺) showed rapid removal of the arsenic in presence of caffeine than those maintained in normal water. Therefore, the caffeine (1,3,7-Trimethylxanthine) as a modulator of Arsenic bioaccumulation in the experimental *Pelecypod* model, *L. corrianus*.

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REFERENCES

- Ravera O, Vido L. Misura del Mn-54. In the population due Unio pictorial, L. (Molluschi Lameellibranchi) Del Lago maggiore. Mem Ist Ital Idrobiol 1961;13:75-84.
- 2. Gaglione P, Ravera O. Manganese-54 concentration in fallout, water and unio mussels of Lake Maggiore, 1960-63. Nature 1964;204:1215-6.
- Ferrington, JW, Goldberg ED, Risebrough RW, Martin JH, Bowen VT. U.S. "Mussel Watch" 1976-1978: An overview of the trace metals, DDE, PCB, hydrocarbon and artificial radionuclide data. Environ Sci Technol 1983;17:490-6.

- Czarnezki JM. Use of the pocketbook mussel, *Lampsilis ventricosa* for monitoring heavy metal pollution in an Ozark stream. Bull Environ Contam Toxicol 1987;38:641-6.
- Doherty FG, Evans DW, Neuhauser EF. An assessment of total and leachable contaminants in zebra mussels (*Dreissena polymorpha*) from Lake Eire. Ecotoxicol Environ Saf 1993;25:328-40.
- Oertel, N. Molluscs as bio monitors of heavy metals in a side-arm system of the River Danube disturbed by engineering activities. Verch Int Veer Limnol 1998;26:2120-4.
- Sures, B, Steiner W, Rydio M, Taraschwski H. Concentrations, if 17 elements in the zebra mussel (*Dreissena polymorpha*) in different tissues of perch. (*Perca fluviatilis*) and in perch intestinal parasites (*Acanthocephalus lucii*) from the sub alpine Lake Mondsee, Austria. Environ Toxiciol Chem 1999;48:2574-9.
- Nayak, L. Heavy metal concentration in two important penaeid prawns from chilka Logoon. Poll Res 1999;18:373-6.
- 9. Bayden CR. Trace metals content and body size in mollusks. Nature 1974;251:311-4.
- Bayden CR. Effect of upon metal content of shellfish. J Mar Biol Assoc UK 1977b;57:675.
- Woolson EA. Bioaccumulation of arsenic. In: Woolson EA, editor. Arsenical pesticides. ACS Symposium Series T. Washington, *DC:* American Chemical Society, 1975.
- Wagemann R, Snow NB, Rosenber DM, Lutz A. Arsenic in sediments, water and aquatic biota from lakes in the vicinity of Yellowknife, Northwest Territories, Canada. Arch Environm Contam Toxicol 1978;7:169-91.
- Spehar RL, Fiandt JT, Anderson RL, DeFoe DI. Comparative toxicity of arsenic compounds and their accumulation in invertebrates and fish. Arch Environm Contam Toxicol 1980;9:53-63.
- Maeada S, Ohki A, Tokuda T, Ohmine M. Transformation of arsenic compounds in a freshwater food chain. Appl Organomet Chem 1990;4:251-4.
- Chen CY, Folt CL. Bioaccumulation and diminution of arsenic and lead in a fresh water food web. Environ Sci Technol 2000;34:3878-84.
- Mason RP, Laporte JM, Andres S. Factors controlling the bioaccumulation of Mercury, methylmercury, arsenic, selenium, and cadmium by freshwater invertebrates and fish. Arch Environ Contam Toxicol 2000;38:283-97.
- Goessler W, Maher W, Irgolic KJ, Kuehnelt D, Schlagenhavfen C, Kaise T. Arsenic compounds in a marine food chain. Fernish J Anal Chem 1997;359:434-7.
- Ochsenkunn Petropulu, Varsamis J, Parissakis G. Speculation of arsenobetaine in marine organisms using a selective bleaching / digestion procedure and hydride generation atomic absorption spectrometry. Anal Chim Acta 1997;337:323-7.
- De Gieter M, Leemakers M, Van Ryssen R, Noyen J, Goeyens L, Baeyens W. Total and toxic arsenic levels in north sea fish. Arch Environ Contam Toxicol 2002;43:406-17.
- Johnson A, Rose M, Inorganic arsenic levels in puget sound fish and shellfish from 303 (d) listed water bodies and other areas. Environmental Assessment Program Olympia, Washington: Washington State Department of Ecology; 2002.
- Kaise T, Ogura M, Nozaki T, Saithoh K, Sakurai T, Matsabara C, *et al.* Biomethylation of arsenic in arsenic Raise freshwater environment. Appl Organomet Chem 1997;11:297-304.
- Meada S, Ohki A, Kusadome K, Kurowa T, Yoshifuku I, Nakas K. Bioaccumulation of arsenic and its fate in a freshwater food chain. Appl Organomet Chem 1992;6:213-19.
- Meada S, Mawatari A, Ohki A, Naka K, Arsenic metabolism in a freshwater food chain: Blue-green alga (*Nostoc* sp) Shrimp (Neocardina *denticulata*) Carp (Cyprinus *carpio*). Appl Organomet Chem 1993;7:467-76.
- 24. Suhendrayatna AO, Maeda S. Biotransformation of arsenate in freshwater food-chain models. Appl Organomet Chem 2001;15:277-84.

Shamsundar and Sureshchandra: Caffeine as a Modulator on Arsenic Bioaccumulation in L. corrianus

- Suhendryatna Ohki A, Nakajima T, Maeda S. Studies on the accumulation and transformation of arsenic in freshwater organisms II. Accumulation and transformation of arsenic compounds by *Tilapia mossambica*. Chemosphere 2002;46:325-31.
- Thomas DJ, Styblo M, Lin S. The cellular metabolism and systemic toxicity of arsenic. Toxicol Appl Pharm 2001;176:127-44.
- Kitchen KT, Ahmad S. Oxidative stress as a possible mode of action for arsenic carcinogenesis. Toxicol Lett 2003;137:3-13.
- Kolayly S, Ocak M, Murat K, Abhasoglu R. A study on doses caffeine bind to a metal ion, elsevier. Food Chem 2004;84:383-88.
- Moore JW, Ramamoorthy S. Heavy metals in natural waters. In: Applied Monitoring and Impact Assessment. New York: Springer Verlag; 1984.
- Muralidharan G, Raja PV. Trace element concentration in the meat of the edible clam, *Marcia resins*, Chemnitz (*Pelecypoda: Veneridae*). Indian J Mar Sci 1997;26:383-85.
- Mitra A, Mitra S, Hazra S, Chaudhari. Heavy metal concentration in India. Coastal Fishes. Res J Chem Environ 2000;4:35.
- Panie S, Bajpai A, Misra SM. Studies on bioaccumulation of selective heavy metals in a tropical ecosystem, Res J Chem Environ 2002;9:67-8.
- Canivet V, Chambon P, Gibert J. Toxicity and bioaccumulation of arsenic and chromium in epigen and hypogean freshwater macro invertebrates, Arch Environ Contam Toxicol 2001;40:345-54.
- Usero J, Regaladoe G, Gracial. Trace metals in the bivalve molluscs *Ruditapes decussatus* and *Ruditapes philippinarum* from the Atlantic coast of Southern Spain: Environ Int1997;23:291-8.
- Goksu MZ, Akar M, Cevik F, Findik O. Bioaccumulation of some heavy metals (Cd, Fe, Zn, Cu) in two Bivalvia Species, Turk. J Vet Anim Sci 2005;29:89-93.
- Pragatheeswaran V. Effect of Zinc, Copper and mercury on Ambassis Commersoni (Cuvier). Ph.D. Thesis, Annamalai University, India, 1987.
- Sayer MD, Reader JP, Morries R. The effect of cadmium concentration on the toxicity of Copper, Lead, Zinc to Yolk Sac of brown trout, *Salmo trout*. L. In soft and acid water J Fish Biol 1989;35:323-32.
- Barber D, Sharma MS. Experimentally induced bioaccumulation and elimination of cadmium in freshwater fishes. Poll Res 1998;17:99-104.
- Senthiloathan S, Balasubramanian T, Venugopal VK. Metal concentration in mussel *Perna vridis* (Bivalvia) and Oyster *Crassostrea madrasensis* (Bivalvia) from some parts of southeast coast of India. Indian J Mar Sci 1998;27:206-10.
- Senthiloathan S, Balasubramanian T. Heavy metal concentration in the oyster Crassostrea *madrasensis* (Bivalve) From the Uppanar. Vellar and Kaduviar estuaries of Southeast coast of India. Indian. J Mar Sci 1998;27:211-16.

- Jeffree RA, Markich SJ, Brown PL. Comparative accumulation of alkaline earth metals by two freshwater mussel species from the Nepean River, Australia: consistencies and a resolved the paradox. Aust J Mar Fre wat Res 1993;44:609-34.
- 42. Metcalfe Smith JL. The influence of species and sex on metal residues in freshwater mussels (Family Unionidae) from the St. Lawrence River, with implications for biomonitoring programs. Environ Toxicol Chem, 1994.
- 43. Mason AZ, Jenkins KD. Metal detoxification in aquatic organisms. In: Tessiex A, Turner DR (Editors). Metal specialty and bioavailability in aquatic systems. New York: John Willey and Sons, 1995. p. 479-589.
- 44. Vesk PA, Byrne M. Metal levels in tissue granules of the freshwater bivalve, *Hyridella depressa* (Unionida) for bio monitoring: The importance of cryoprescrvation. The Science of the Total. Envit 1999;225:219-29.
- 45. Byrne M. Calcium concretions in the interstitial tissues of the Australian freshwater mussel *Hyridella depressa* (*Hyriidae*). In: Harper EM, Taylor JD, Crame JA, editors. The Evolutionary Biology of the Bivalvia Geological Society. Vol. 177. London: Special Publication; 2000. p. 329-37.
- Leoty C, Hatchet- Cadiou C, Talon S, Choisy S, Hleihel W. Caffeine stimulates the reserve mode of the Na /Ca2+ exchanges in ferret ventricular muscles, Acta Physiol Scand 2001;172:27-37.
- 47. Sugiyama K, Noda Y, He P. Suppressive effect of caffeine on hepatitis and apoptosis induced by tumour necrosis factor- L but not by the anti-Fas antibody in mice. Biosci Biotechnol Biochem 2001;65:674-7.
- Abbasoglu R, Kucwk M, Kolayl S, Ocak M. Theoretical investigation theophylline. In proceeding of the Int. Congress on the chemistry of Natural Product S. Trabzon, Turkiya, 2002.
- Hoska S, Kawa S, Aoki V, Tanoka E, Yoshizawak K, Karasaw Y, *et al.* Hepatocarcinogensis inhibition by caffeine in ACI rats treated with 2- acetyl aminoflurence. Food Chem Toxicol 2001;39:557-61.
- Lu T, Liu J, LeCluyse EL, Zhou YS, Cheng ML, Waalkes MP. Applications of cDNA microarray to the study of arsenic induced liver diseases in the population of Guizhou, China. Toxicol Sci 2001;59:185-92.
- Hove-Madsen L, Llach A, Tort L. Quantification of calcium release from the sarcoplasmic reticulum in rainbow trout atrial myocytes. Pflugers Arch 1999;438:545-52.

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